

# Review Article: Bioactive Compounds and Antischistosomal Activity of Dolichos Species: A Review

Nasiru Malan Musa<sup>1,\*</sup>, Muhammad Sani Sallau<sup>2</sup>, Adebayo Ojo Oyewale<sup>2</sup>, Tijjani Ali<sup>3</sup>

<sup>1</sup>Department of Chemistry, College of Natural and Applied Sciences, Al-Qalam University Katsina, Katsina State, Nigeria

<sup>2</sup>Department of Chemistry, Faculty of Physical Sciences, Ahmadu Bello University Zaria, Kaduna State, Nigeria

<sup>3</sup>Department of Chemistry, Faculty of Physical Sciences, Federal University Dutsin-Ma, Katsina State, Nigeria



**Citation:** N.M. Musa\*, M.S. Sallau, A.O. Oyewale, T. Ali, Bioactive Compounds and Antischistosomal Activity of Dolichos Species: A Review. *J. Chem. Rev.*, 2023, 5(4), 416-438.

<https://doi.org/10.48309/JCR.2023.407571.1233>



#### Article info:

Received: 17 July 2023

Accepted: 15 August 2023

Available Online: 21 August 2023

ID: JCR-2307-1233

Checked for Plagiarism: Yes

Language Editor: Dr. Fatimah

Ramezani

Editor who Approved Publication:

Prof. Dr. Ghasem Rezanejade Bardajee

#### Keywords:

Dolichos species, Isolation, Antischistosomal activity, Medicinal plants, Bioactive compounds

## ABSTRACT

This review focuses on the isolated bioactive compounds and antischistosomal activity of Dolichos species, particularly Dolichos biflorus and Dolichos lablab. Medicinal plants have been used since ancient times and continue to play a significant role in traditional medicine and drug discovery. Dolichos species are known for their therapeutic potential and have been traditionally used in Nigeria. The review presented the chemical compounds isolated from Dolichos species, including triterpene bisdesmosides and lectins. Compounds with antischistosomal activity were also explored. The findings highlight the importance of Dolichos species as potential sources of bioactive compounds for the development of novel therapeutic agents.



**Nasiru Malan Musa:** He is a Ph.D. student in Organic Chemistry Ahmadu Bello University Zaria, Kaduna State and a lecturer in the Department of Chemistry, College of Natural and Applied Sciences, Al-Qalam University Katsina, Katsina State, Nigeria.

\*Corresponding Author: Nasiru Malan Musa ([nmmusabr@gmail.com](mailto:nmmusabr@gmail.com))



**Muhammad Sani Sallau:** He is a co-author and professor in the Department of Chemistry, Faculty of Physical Sciences, Ahmadu Bello University, Zaria Kaduna State, Nigeria. He is skilled in Curriculum Design, Matlab, Simulation.



**Adebayo Ojo Oyewale:** He is a co-author and professor in the Department of Chemistry, Faculty of Physical Sciences, Ahmadu Bello University Zaria, Kaduna State Nigeria. He specialized in synthesis, isolation, and characterization of organic molecules using 1D and 2D NMR spectroscopy.



**Tijjani Ali:** He is a co-author and associate professor in the Department of Chemistry, Federal University Dutsin-Ma, Katsina State, Nigeria. His research interest mainly focuses on plant-derived antimicrobial naturally occurring molecules. He specialized on isolation, characterization, and pharmacological studies of phytochemicals.

---

## Content

1. Introduction
2. Results and Discussion
  - 2.1 Compounds isolated from Dolichos species
  - 2.2. Compounds isolated from Dolichos family
  - 2.3. Some compounds with antischistosomal activity
3. Conclusion

## 1. Introduction

Since the origin of medicine, natural products, particularly those produced from plants, have been used to help mankind maintain human health [1].

Traditional medicine has existed since the beginning of time and it has been widely acknowledged and used by people throughout history [2]. Plants have been utilized as a source of medicine since the dawn of humanity [3, 4]. For many years, plant-derived therapeutic medicines have piqued the interest of experts all over the world due to their low side effects and good impacts on human health [5]. Plant's use in medicine dates back to the Greek physician Dioscorides, who published *De Materia Medica* in 28 A.D., which comprised 600 medicinal plants and was the main work on pharmacology until the Renaissance [6, 7]. Willow tree leaves of the plant may have been prescribed by Hippocrates to treat fever [8, 9]. Salicin, an anti-inflammatory and pain-relieving compound, was initially isolated from the white willow tree, and then synthesized to become a popular over-the-counter medication [10, 11]. Plants having a long history of ethnomedicinal uses can be a valuable source of substances for the treatment of various ailments and infectious diseases in the pharmaceutical field [12].

Medicinal plants are thought to be a reservoir of various bioactive chemicals with various medicinal qualities [13]. Anti-inflammatory, antiviral, anticancer, antimalarial, and analgesic activities are among the many therapeutic benefits linked with medicinal plants [14, 15]. According to the World Health Organization (WHO), diverse medicinal plants can be used to make a range of pharmaceuticals [16] and around 80 % of the world's developing population relies on traditional medicines for their primary health care [1, 17].

Plants have a natural advantage of producing pharmaceutical chemicals [18], and their characterization has led to the identification of new, low-cost medications with excellent therapeutic potential [19, 20]. Medicinal plants play an important role in rural livelihoods [21]. Many people are active in collecting and trading medicinal plants [22], in addition to traditional

healers who practice herbal medicine [23]. The result is an increased demand in both local and international markets as well as bio-prospecting activities searching for sources of new drugs [24, 25].

According to the World Health Organization (WHO), the activity of medicinal plants are not only limited to boosting flora, but also recognized for their medicinal value [26-29]. Antimicrobial, antioxidant, anticancer, hypolipidemic, cardiovascular, central nervous, respiratory, immunological, anti-inflammatory, analgesic antipyretic, and many other pharmacological actions were found in the plant [25, 30, 31].

Between 1983 and 1994, chemicals derived from natural products or semi-synthetic drugs derived from natural sources accounted for 78 percent of new drugs approved by the US Food and Drug Administration (FDA) [14, 32, 33]. According to a poll, the public's use of medicinal plants increased from 3 % in 1993 to over 37 % in 1998 [34, 35]. The low cost of herbal drugs, which endears them to the poor masses of the developing world, the 'green' movement in the developed world, which campaigns on the inherent safety and desirability of natural products; and the individualistic philosophy of western society, that further encourages self-medication, with many people preferring to treat themselves with phytomedicines, have all facilitated this shift to herbal drugs [10, 36, 37]. *Dolichos* is a genus of Papilionaceae family. In Nigeria, there are about 8 kinds, with *Dolichos biflorus* (*D. biflorus*) and *Dolichos lablab* (*D. lablab*) being the most widely cultivated and used [38]. Since antiquity, the plant has been far more popular in Nigeria and India than in any other country [39]. As a result, the purpose of this review was to describe the chemical compounds and antischistosomal activity of some *Dolichos* species currently reported.

## 2. Results and Discussion

### 2.1 Compounds isolated from *Dolichos* species

Mono and oligosaccharides (7.3%) were isolated from the endosperm of *Dolichos pachyrhizus* [40]. Fructose (1), galactose (2),

glucose (3), sucrose (4), and raffinose (5) were recognized [41]. Six novel oleanane type triterpene bisdesmosides were recovered from the glycoside combination of *Dolichos pachyrhizus* seeds, together with chikusetsu saponin IV. *Dolichos pachyrhizus* var. *lignosus* (field bean) and *Dolichos pachyrhizus* var. *typicus* seeds were used to isolate two lectins (Pachyrhizus bean) [20]. Both lectins appeared to be formed up of four identical subunits and had a molecular weight of 60,000 (apparent molecular weight 15,000). The lectins' carbohydrate content was primarily fructose (2-5 moles per mol of protein) (1), mannose (5-8 mol per mol of protein) (6) and *N*-acetyl glucosamine (1-2 moles per mol of protein) (7). Both lectins have a comparable amino acid composition and a similar tryptic peptide map [42].

Both lectins have only alanine and serine as N and C-terminal amino acids. Low levels of bound metals such as manganese, magnesium, and calcium were identified in the lectins. The lectins' near-ultraviolet circular dichroism spectra were similar to sainfoin's [43]. Tyrosine and tryptophan residues were engaged in sugar binding, according to circular dichroism studies [44]. *Pachyrhizus pachyrhizus* contains a total of 262 volatile chemicals, volatile terpenes, and terpenoids, as well as their derivatives, dominated the volatile components, accounting for 46 % of all identified molecules [45]. The detected compounds were separated into 12 classes, namely alcohols (28), aldehydes (10), ketones (19), esters (46), acids (7), oxygen heterocycles (1), pyrazines (5), thiazoles (4), hydrocarbons (57), terpenes and terpenoids (59), phenols (5), and miscellaneous compounds [46]. The most common individual compounds were Isopentyl alcohol (8), 3,7,11-trimethylhentriacontane (9), (*E*)-2-octene (10), 7, 11, 17, 21-tetramethylhentriacontane/ 7, 11, 17, 25-tetramethylhentriacontane (11), 6-methyldotriacontane (12), norbornene (13), pentanol (14), 4-methylthiazole (15), 5,9,13-trimethylnonacosane (16), methylbutyrate (17), isopentylformate (18), 13,17-dimethylnonacosane (19), 13-methylhentriacontane (20), 9-

methylhentriacontane (21), 7-methylhentriacontane (22), heptanal (23), 5-methylhentriacontane (24), 3, 11, 19-trimethylhentriacontane (25), and 3, 7-dimethylhentriacontane (26) [4]. Luteolin (27), luteolin-4'-*O*-*D*-glucopyranoside (28), and luteolin-7-*O*- $\beta$ -*D*-glucopyranoside (29) were found as flavonoids isolated from the flower of *Dolichos pachyrhizus* (Table 1) [35]. The rotenoid content of *Pachyrhizus purpureus* plant sections was investigated. The roots had the highest amount, while the seeds had the lowest [47].

## 2.2. Compounds isolated from Dolichos family

Studies have been carried out on some *Dolichos* species to identify the presence of several constituents. Having extensively studied the phytochemistry of *Dolichos*, different research groups have identified the following compounds from the root extract of *Neorautanenia mitis* [7]: Neodulene (30) [48], Neodulin (31), Ferullic acid (32) [49], Pachyrhizine (33), Neotenone (34), 12a-hydroxydolioneon (35), Dolineon (36), Dehydroneotenone (37), [50], Ambonane (38) [51], Stigmasterole (39) [52], 7-methoxy-3-(6-methoxybenzo[*d*][1,3]dioxol-5-yl)chroman-4-one (40) [53], (-)-2-isopentenyl-3-hydroxy-8-9-methylenedioxypterocarpan (41) [54], Nepseudin (42) [55], Neorautenol (43) [56, 57], Isoneorautenol (44) [58], (-)-2-hydroxypterocarpin (45) [59], Rotenone (46), 12a-hydroxyrotenone (47) [60-61], Rautandiol A (48), and Rautandiol B (49) (Table 2) [49, 62].

## 2.3. Some compounds with antischistosomal activity

Studies were carried out from different plant species and assessed for the antischistosomal activity. In a study conducted by [63], the following compound were isolated and tested for their antimicrobial and antischistosomal activities from the leaves extracts of *E. camaldulensis*; Gallic acid (50), Taxifolin (51), Methyl gallate (52), Quercetin (53), Luteolin



Table 1. Continued


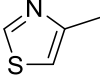
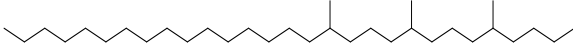
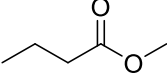
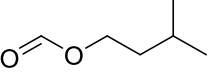
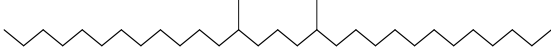
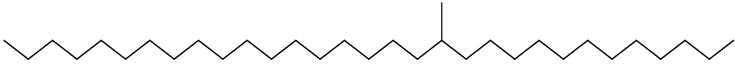
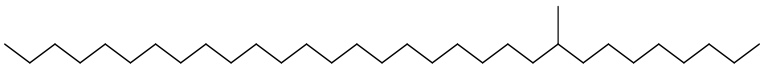
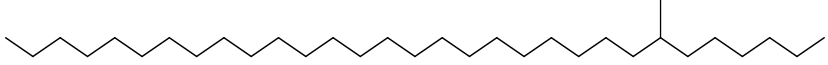
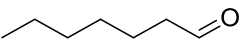
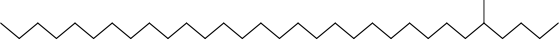
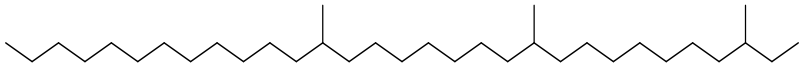
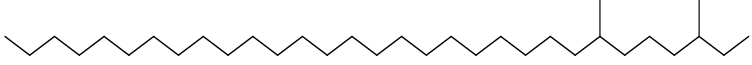
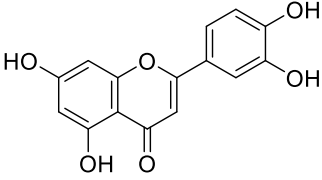
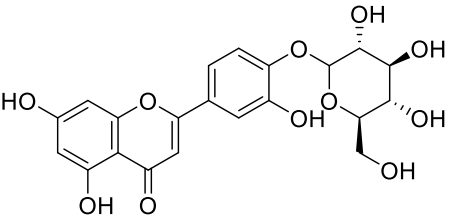
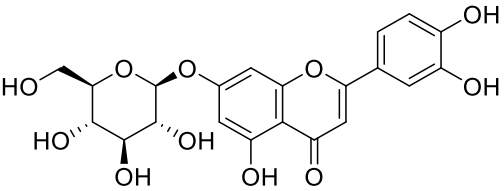
<p><i>Pachyrhizus pachyrhizus</i> (seeds)</p>	<div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(14)</p> </div> <div style="text-align: center;">  <p>(15)</p> </div> </div> <div style="display: flex; justify-content: space-around; margin-top: 20px;"> <div style="text-align: center;">  <p>(16)</p> </div> <div style="text-align: center;">  <p>(17)</p> </div> </div> <div style="display: flex; justify-content: space-around; margin-top: 20px;"> <div style="text-align: center;">  <p>(18)</p> </div> <div style="text-align: center;">  <p>(19)</p> </div> </div> <div style="text-align: center; margin-top: 20px;">  <p>(20)</p> </div> <div style="text-align: center; margin-top: 20px;">  <p>(21)</p> </div> <div style="text-align: center; margin-top: 20px;">  <p>(22)</p> </div> <div style="display: flex; justify-content: space-around; margin-top: 20px;"> <div style="text-align: center;">  <p>(23)</p> </div> <div style="text-align: center;">  <p>(24)</p> </div> </div> <div style="text-align: center; margin-top: 20px;">  <p>(25)</p> </div> <div style="text-align: center; margin-top: 20px;">  <p>(26)</p> </div>	[4]
<p><i>Dolichos pachyrhizus</i> (flowers)</p>	<div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(27)</p> </div> <div style="text-align: center;">  <p>(28)</p> </div> </div> <div style="text-align: center; margin-top: 20px;">  <p>(29)</p> </div>	[35];[47]

Table 2. Compounds isolated from Dolichos family

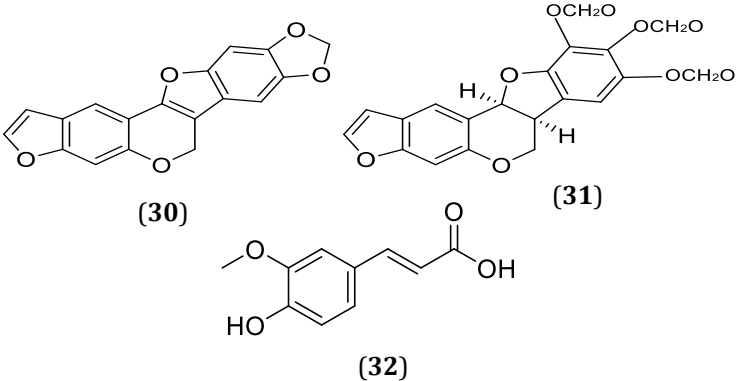
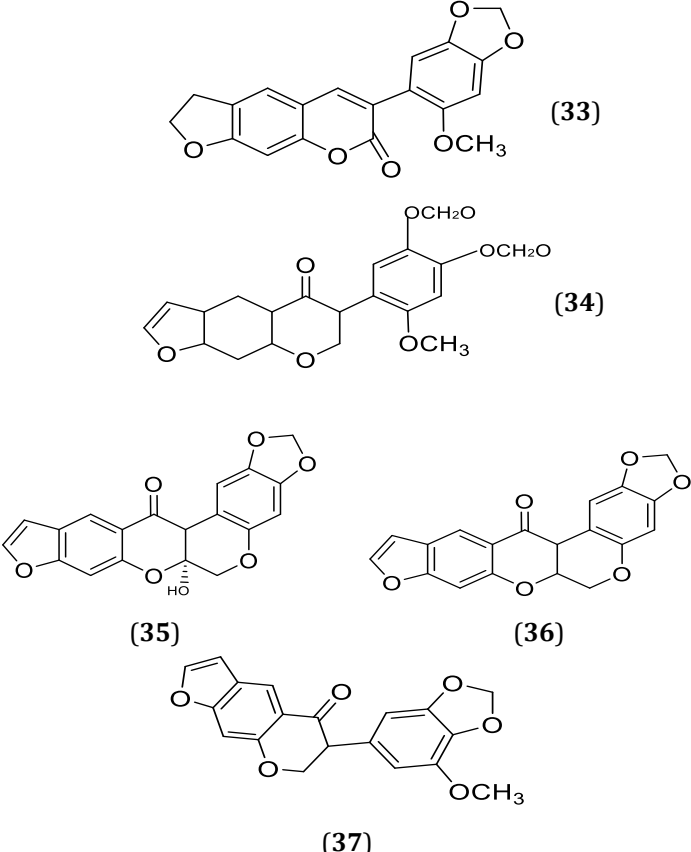
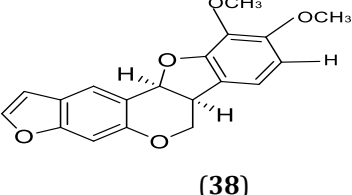
Botanical name	Name of compound	Reference
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(30) (31) (32)</p>	[49]
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(33) (34) (35) (36) (37)</p>	[50]
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(38)</p>	[51]

Table 2. Continued

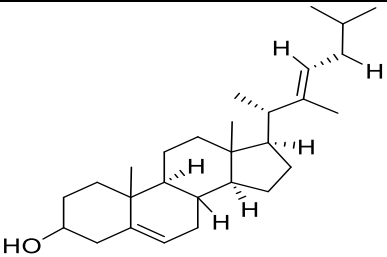
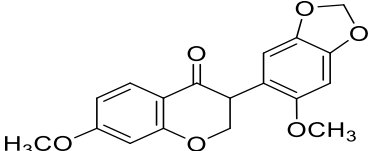
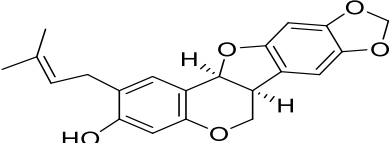
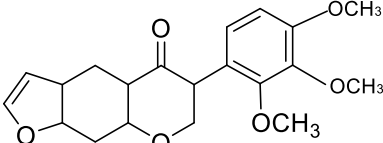
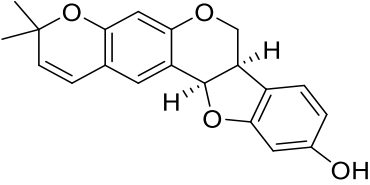
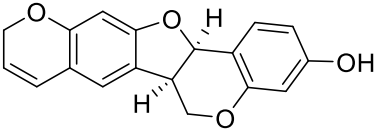
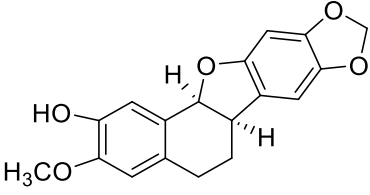
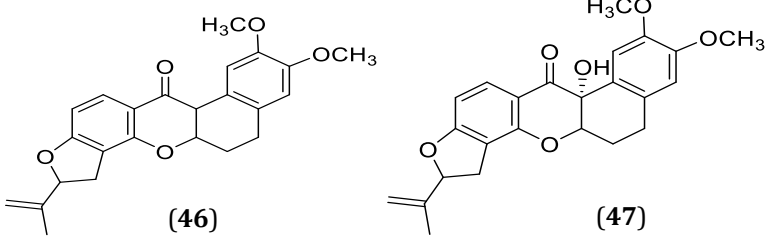
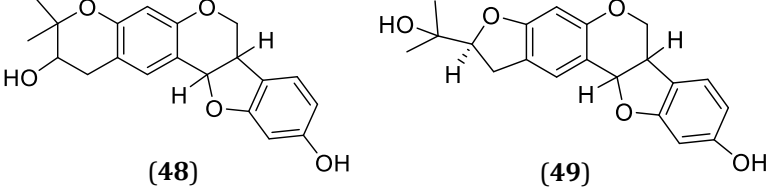
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(39)</p>	<p>[52]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(40)</p>	<p>[53]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(41)</p>	<p>[54]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(42)</p>	<p>[55]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(43)</p>	<p>[56-57]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(44)</p>	<p>[58]</p>
<p><i>Neorautanenia mitis</i> (root)</p>	 <p>(45)</p>	<p>[59]</p>



Table 2. Continued

<i>Neorautanenia mitis</i> (root)	 <p style="text-align: center;">(46)                      (47)</p>	[60-61]
<i>Neorautanenia mitis</i> (root)	 <p style="text-align: center;">(48)                      (49)</p>	[49]

(54), and Hesperidin (55). A triterpenoid Asparagalin (56) was also isolated from *Asparagus stipularis* an African medicinal plant [64]. *In vitro* Antischistosomal activities of the extracts and compounds from *Solidago microglossa* DC (Asteraceae), *Aristolochia cymbifera* Mart., and Zucc. (Aristolochiaceae) were assessed and revealed the effectiveness of the following compounds against schistosomal larva; Bauerenol (57), alpha-amyrin (58), Spinasterol (59), Populifolic acid (60), Cubebin (61), 2-oxopopulifolic acid methyl ester (62), and 2-oxo populifolic acid (63) [65].

In a research conducted by [66], reported the isolation of thirty nine (39) compounds with antischistosomal activities including the terpenoids: Rotundifolone (64) [66-70], Limonine epoxide (65), and Carvacryl acetate (66) [71], Artemisinin (67) [66,69,72], Artemether (68), and Artesunate (69) [67-69,73-74], Dihydroartemisinin (70) [66,69,75-

76], Nerolidol (71) [77], Budlein-A (72), Dihydrobudlein-A (73), and Tetrahydrobudlein-A (74) [78], Phytol (75) [79], Betulin (76) [80], Triphenylphosphonium salt of betulin (77) and (78) [81], Balsaminol F (79), and Karavilagenin C (80) [82]; the alkaloids: Piplartine (81) [83], Piperamide (82) [84], Epiisopiloturine (83) [85], Sanguinarine (84), Solamargine (85) and Solasanine (86) [86], Mefloquine (87) [87-88], and the phenolics: Plumbagin (88) [89-90], b-Lapachone (89) [91], Quarcetin-3-O-b-d-glucoside (90) Quarcetin-3-O-b-d-rhamnoside (91) [92], Kaempferol (92) [93], Sativan (93) [94], Hesperidin (94) [95], Licarin A (95) [96], Curcumin (96) [97-100], Aspidin a phloroglucinol derivative (97), Flavaspodic acid (98), Methylene-bis-aspidinol (99), and Desaspidin (100) (Table 3) [101].

**Table 3.** Some compounds with antischistosomal activity

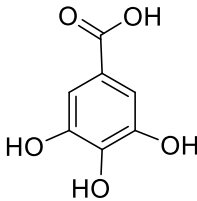
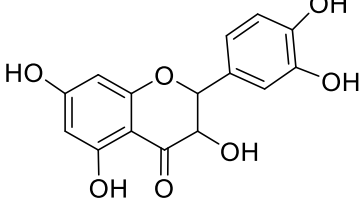
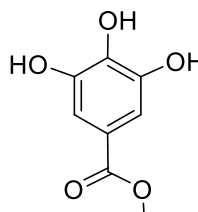
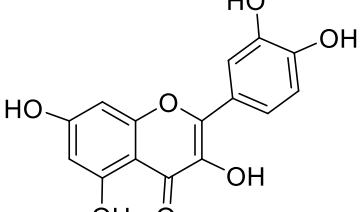
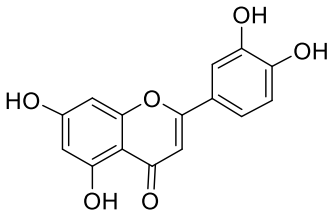
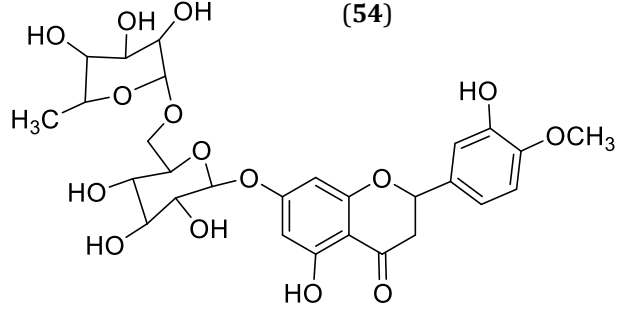
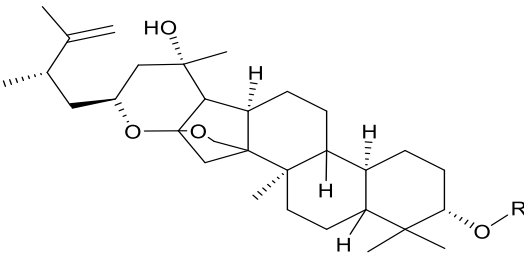
Botanical name	Name of compound	Reference	
<i>E. camaldulensis</i> (leaves)	 <p>(50)</p>	[63]	
	 <p>(51)</p>		
	 <p>(52)</p>		 <p>(53)</p>
	 <p>(54)</p>		
	 <p>(55)</p>		
<i>Asparagus stipularis</i>	 <p>Asparagalin (56)</p>	[64]	

Table 3. Continued

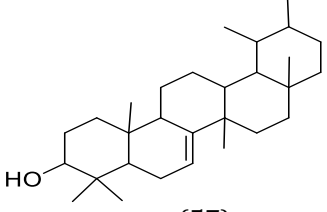
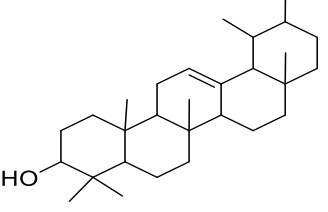
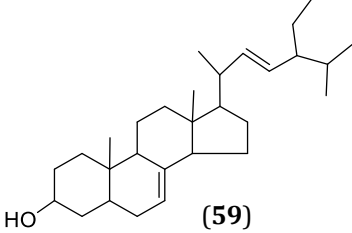
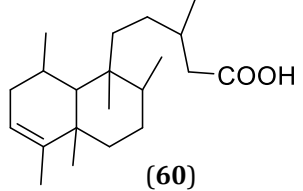
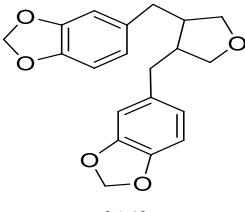
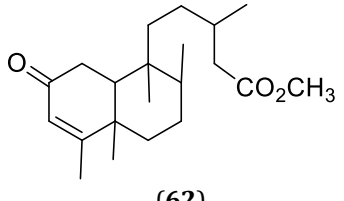
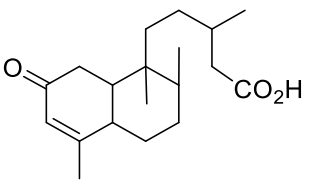
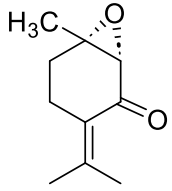
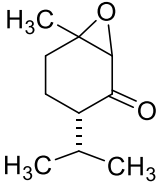
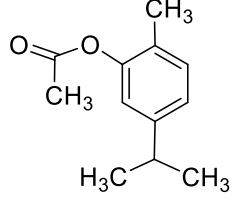
<p><i>Solidago microglossa</i> and <i>Aristolochia</i> <i>cymbifera</i> (extract)</p>	<div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(57)</p> </div> <div style="text-align: center;">  <p>(58)</p> </div> </div> <div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(59)</p> </div> <div style="text-align: center;">  <p>(60)</p> </div> </div> <div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(61)</p> </div> <div style="text-align: center;">  <p>(62)</p> </div> </div> <div style="text-align: center; margin-top: 20px;">  <p>(63)</p> </div>	<p>[65]</p>
<p><i>Mentha villosa</i></p>	<div style="text-align: center;">  <p>(64)</p> </div>	<p>[66-70]</p>
<p>Derived from many plants</p>	<div style="display: flex; justify-content: space-around;"> <div style="text-align: center;">  <p>(65)</p> </div> <div style="text-align: center;">  <p>(66)</p> </div> </div>	<p>[71]</p>

Table 3. Continued

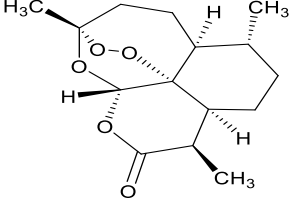
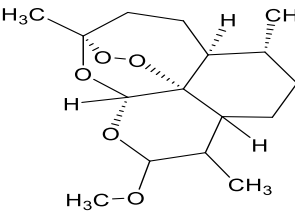
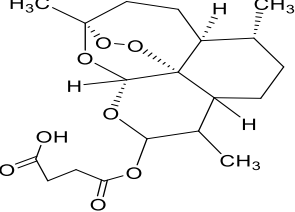
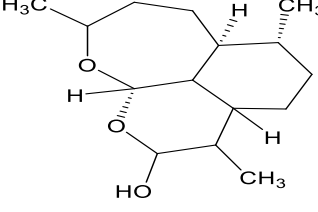
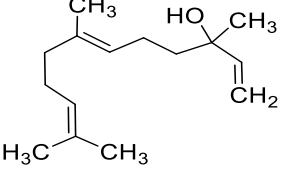
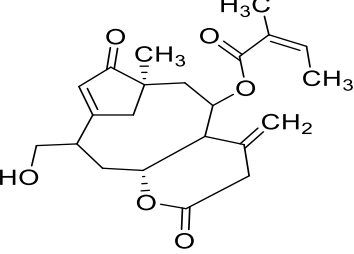
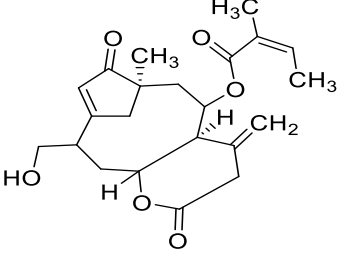
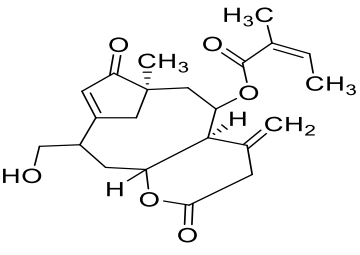
<i>Artemisia annua</i> L	 <p>(67)</p>	[66;69;72]
Derived from artemisinin	 <p>(68)</p>  <p>(69)</p>	[67-69] [73-74]
Derived from artemisinin	 <p>(70)</p>	[66];[69] [75-76]
Derived from many plants	 <p>(71)</p>	[77]
<i>Artemisia annua</i> L	 <p>(72)</p>  <p>(73)</p>  <p>(74)</p>	[78]

Table 3. Continued

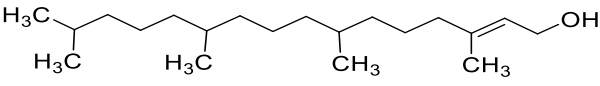
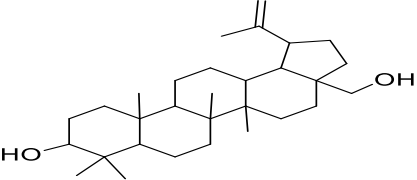
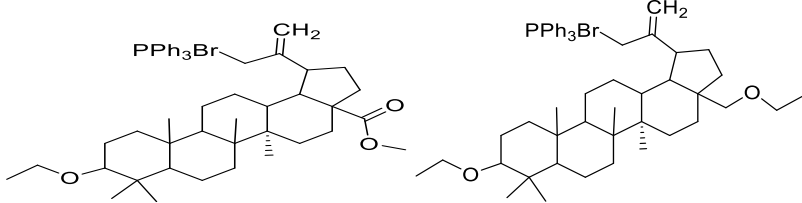
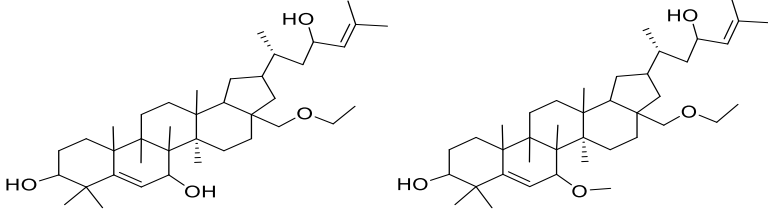
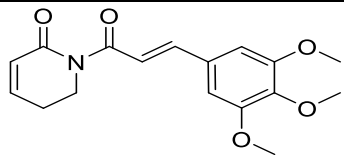
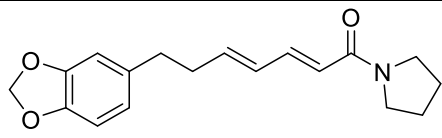
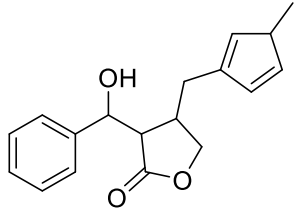
Derived from many plants	 <p style="text-align: center;">(75)</p>	[79]
<i>Schefflera vinosa</i>	 <p style="text-align: center;">(76)</p>	[80]
Derived from botulin	 <p style="text-align: center;">(77) (78)</p>	[81]
<i>Momordica balsamina</i> L	 <p style="text-align: center;">(79) (80)</p>	[82]
<i>Piper tuberculatum</i>	 <p style="text-align: center;">(81)</p>	[83]
<i>Piper amalago</i> L	 <p style="text-align: center;">(82)</p>	[84]
<i>Pilocarpus microphyllus</i> Stapf	 <p style="text-align: center;">(83)</p>	[85]

Table 3. Continued

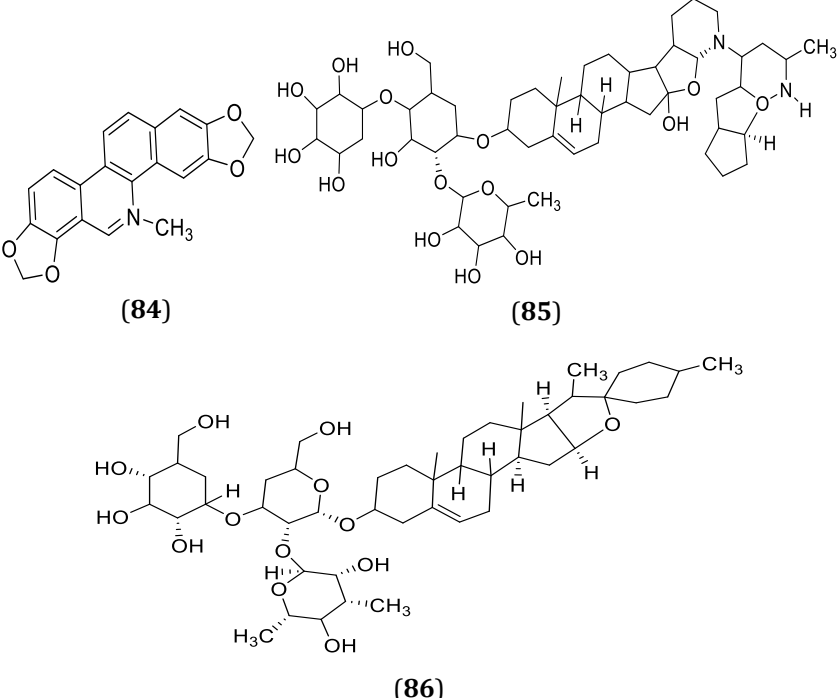
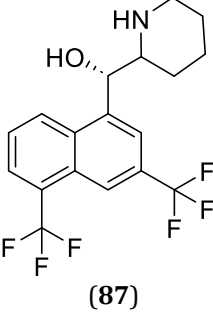
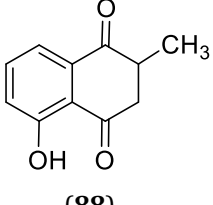
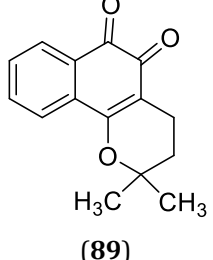
Sanguinaria spp	 <p>(84) (85) (86)</p>	[86]
Analogue of quinine	 <p>(87)</p>	[87-88]
Plumbago spp	 <p>(88)</p>	[89-90]
Derived from lapachol	 <p>(89)</p>	[91]

Table 3. Continued

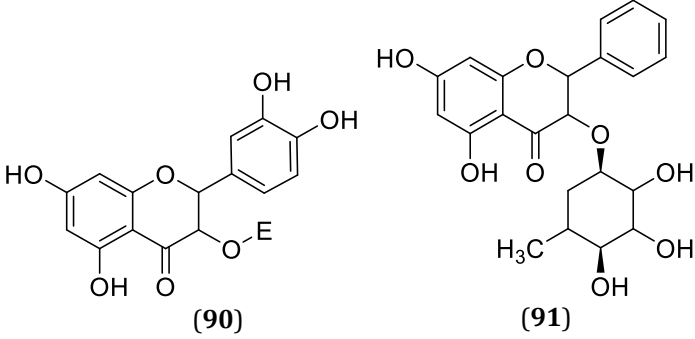
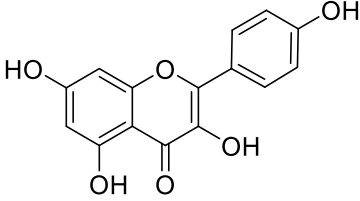
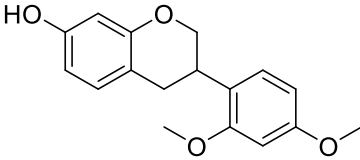
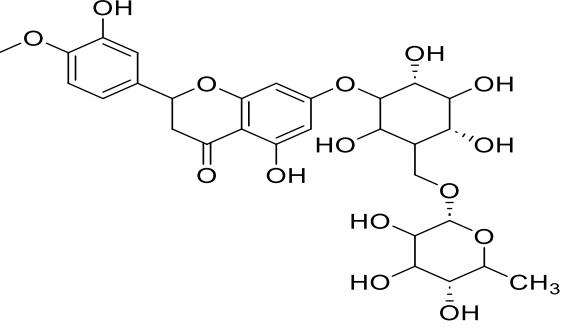
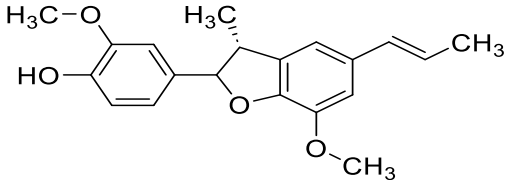
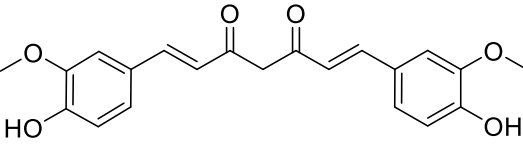
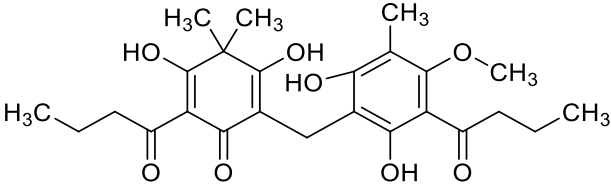
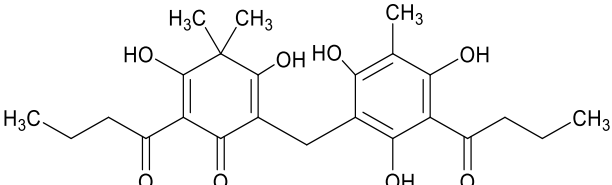
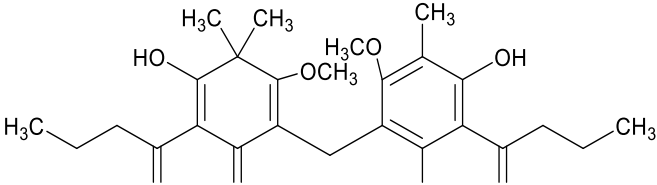
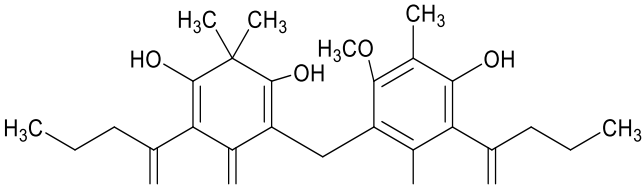
<i>Roupala montana</i>	 <p>(90) (91)</p>	[92]
<i>Styrax pohlii</i> Pohl	 <p>(92)</p>	[93]
<i>Astragalus englerianus</i> Ulbr.	 <p>(93)</p>	[94]
Citrus fruits	 <p>(94)</p>	[95]
Many plants.	 <p>(95)</p>	[96]
<i>Curcuma longa</i> L.	 <p>(96)</p>	[95]; [97-100]

Table 3. Continued

Dryopteris genus	 <p style="text-align: center;">(97)</p>	[101]
	 <p style="text-align: center;">(98)</p>	
	 <p style="text-align: center;">(99)</p>	
	 <p style="text-align: center;">(100)</p>	

### 3. Conclusion

The isolation of compounds from *Dolichos* species provides insights into their chemical composition and supports the exploration of their pharmacological activities. The identification of mono and oligosaccharides, triterpene bisdesmosides, and lectins from *Dolichos* species contributes to the understanding of their therapeutic potential. Further research is needed to fully elucidate the mechanisms of action and therapeutic potential of the isolated compounds. In addition, more studies are required to explore other *Dolichos*

species and their bioactive constituents. The continued investigation of *Dolichos* species and their bioactive compounds may lead to the development of new drugs for the treatment of various ailments, including schistosomiasis.

### Acknowledgment

The authors would like to present their gratitude to the laboratory technologist from Ahmadu Bello University Zaria, Federal University, Dutsin-Ma, Katsina State, and Al-Qalam University Katsina for their contribution



in one way or the other during the conduct of the research.

### Conflict of interest

The authors declare that they have no competing interests.

### Orcid:

Nasiru Malan Musa

<https://orcid.org/0000-0002-4113-7228>

Muhammad Sani Sallau

<https://orcid.org/0000-0002-4113-7228>

Adebayo Ojo Oyewale

<https://orcid.org/0000-0002-4113-7228>

Tijjani Ali

<https://orcid.org/0000-0002-4113-7228>

### References

- [1]. M.G. Rasul, Extraction, isolation and characterization of natural products from medicinal plants, *International Journal of Basic Sciences and Applied Computing (IJBSAC)*, **2018**, *2*, 1-6. [Crossref], [Google Scholar]
- [2]. J. Sathya, F.G. Shoba, A study on the phytochemistry and antioxidant effect of methanolic extract of *Citrullus lanatus* seed. *Asian Journal of Plant Science and Research*, **2014**, *4*, 35-40. [Google Scholar], [Publisher]
- [3]. A.J. Tajudeen, A. Tukur, N.M. Musa, R.M. Obansa, M. Isma'il, Phytochemical and gas chromatography-mass spectrometry (GC-MS) analysis of ethyl acetate root extract of *Indigofera diphylla*, *Advanced Journal of Chemistry, Section B*, **2023**, *5*, 173-183. [Crossref], [Google Scholar], [Publisher]
- [4]. S. Abirami, K. Nishanthini, M. Poonkothai, Antimicrobial activity and phytochemical screening of the leaf extracts of *Eucalyptus globulus*. *International Journal of Current Pharmaceutical Research*, **2017**, *9*, 85-89. [Crossref], [Google Scholar], [Publisher]
- [5]. A. Tukur, N.M. Musa, H.A. Bello, N.A. Sani, Determination of the phytochemical constituents and antifungal properties of *Annona senegalensis* leaves (African custard apple), *ChemSearch Journal*, **2020**, *11*, 16-24. [Crossref], [Google Scholar], [Publisher]
- [6]. A. Tukur, N.M. Musa, M. Isma'il, M. Yusuf, J.A. Tajuddeen, Phytochemical screening and evaluation of the antioxidant potentials of the stem bark extracts of *Erythrophleum africanum* (African blackwood), *Advanced Journal of Chemistry, Section B*, **2022**, *4*, 202-208. [Crossref], [Google Scholar], [Publisher]
- [7]. C.J. Dawurung, G.G. Jurbe, J.G. Usman, I.L. Elisha, L.H. Lombin, S.G. Pyne, Antidiarrheal activity of some selected Nigerian plants used in traditional medicine, *Pharmacognosy Research*, **2019**, *11*, 371-777. [Crossref], [Google Scholar], [Publisher]
- [8]. H.Y. Kim, B.H. Moon, H.J. Lee, D.H. Choi, Flavonol glycosides from the leaves of *Eucommia ulmoides* O. with glycation inhibitory activity, *Journal of Ethnopharmacology*, **2004**, *93*, 227-230. [Crossref], [Google Scholar], [Publisher]
- [9]. M. Thapliyal, A. Bisht, A. Singh, Isolation of antibacterial protein/peptide from *Ficus glomerata* leaf, *International Journal of Current Pharmaceutical Research*, **2016**, *8*, 24-27. [Crossref], [Google Scholar], [Publisher]
- [10]. P.J. Houghton, The role of plants in traditional medicine and current therapy, *The Journal of Alternative Complementary Medicine*, **1995**, *1*, 131-143. [Crossref], [Google Scholar], [Publisher]
- [11]. R.A. Moreau, B.D. Whitaker, K.B. Hicks, Phytosterols, phytostanols, and their conjugates in foods: Structural diversity, quantitative analysis, and health-promoting uses, *Progress in Lipid Research*, **2002**, *41*, 457-500. [Crossref], [Google Scholar], [Publisher]
- [12]. a) S. Kumar, A.L. Sheba, A study on phytochemicals, antimicrobial, and synergistic antimicrobial activities of *Hibiscus sabdariffa*, *Asian Journal of Pharmaceutical and Clinical Research*, **2019**, *12*, 198-201. [Crossref], [Google Scholar], [Publisher] b) A.S. Zaek, B.A. Benhamed, M.A. Al shahomy, R. kamour, A.

- Eshames, Comparative study of pharmaceutical content of three different cardio vascular system drugs marketed in Tripoli- Libya, *Progress in Chemical and Biochemical Research*, **2019**, *2*, 6-12. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [13]. L.J. Nohynek, H.L. Alakomi, M.P. Kahkonen, M. Heinonen, I.M. Helander, K.M. Oksman-Caldentey, R.H. Puupponen-Pimia, Berry phenolics: antimicrobial properties and mechanisms of action against severe human pathogens, *Nutrition and cancer*, **2006**, *54*, 18-32, [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [14]. V. Duraipandiyar, M. Ayyanar, S. Ignacimuthu, Antimicrobial activity of some ethnomedicinal plants used by Paliyar tribe from Tamil Nadu, India, *BMC Complementary Medicine and Therapies*, **2006**, *6*, 35-41. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [15]. A.E. Al-Snafi, Therapeutic properties of medicinal plants: a review of their antibacterial activity (part 1), *International Journal of Pharmacology and Toxicology*, **2015**, *6*, 137-158. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [16]. O. Odeja, C.E. Ogwuche, E.E. Elemike, G. Obi, Phytochemical screening, antioxidant and antimicrobial activities of *Acalypha ciliata* plant, *Clinical Phytoscience*, **2016**, *2*, 12. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [17]. a) E.S. Abdel-Hameed, H.A. EL-Nahas, A.S. Abo-Sedera, Antischistosomal and antimicrobial activities of some Egyptian plant species, *Pharmaceutical Biology*, **2008**, *46*, 626-633. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)] b) I. Ogbuwu, B.N. Ijere, Trace metal concentrations in herbal medicine sole in abalaliki metropolis, *Asian Journal of Green Chemistry*, **2022**, *6*, 320-326. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [18]. A. Altemimi, N. Lakhssassi, A. Baharlouei, D.G. Watson, D.A. Lightfoot, Phytochemicals: Extraction, isolation and identification of bioactive compounds from plant extracts. A plant review, *Plants*, **2017**, *6*, 42. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [19]. T.C. Birdsall, G.S. Kelly, Berberine: Therapeutic potential of an alkaloid found in several medicinal plants, *Alternative Medicine Review*, **1997**, *2*, 94-103. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [20]. S.K. Yang, K. Yusoff, C.W. Mai, W.M. Lim, W.S. Yap, S.H.E. Lim, K.S. Lai, Additivity vs synergism: investigation of the additive interaction of cinnamon bark oil and meropenem in combinatory therapy, *Molucule*, **2017**, *22*, 1733. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [21]. K. Poole, *Pseudomonas aeruginosa*: resistance to the max, *Frontiers in Microbiology*, **2011**, *2*, 1-13. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [22]. a) O.P. Ajagbonna, H.O. Oshagbemi, B.R. Olorede, Antibacterial properties of calyx, stem bark and root of *Hibiscus sabdariffa*, *Nigerian Journal of Natural Products and Medicine*, **2001**, *5*, 54-55. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)] b) O. Iyun, J. Ahmed, S.S. Muhammad, I. Hamisu, GC-MS analysis of methanol extract of *Strychnos innocua* (Delile) root bark, *Advanced Journal of Chemistry, Section A*, **2022**, *5*, 104-117. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [23]. J. Rohini, N.V. Mansoureh, S.R.A. Fouad, M.S. Rabeta, A.M. Shah, Preliminary screening on wound healing potential of *Ocimum tenuiflorum* L. using *in vitro* assays, *Food Research*, **2019**, *3*, 258-264. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [24]. G.V.P. Kumar, S.N. Subrahmanyam, Phytochemical analysis, in-vitro screening for antimicrobial and anthelmintic activity of combined hydroalcoholic seed extracts of four selected folklore Indian medicinal plants, *Der Pharmacia Lettre*, **2013**, *5*, 168-176. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [25]. A. Tukur, N.M. Musa, J. T. Abdullahi, J.D. Habila, M.O. Bamidele, Phytochemical screening and the effect of methanolic leaves extract of *Senna mimosoides* on inflammatory Stimulus-Induced leukocyte mobilization (in-vivo), *Advanced Journal of Chemistry, Section B*, **2022**,

- 4, 29-38. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [26]. M. Liu, X.Q. Li, C. Weber, C.Y. Lee, J. Brown, R.H. Liu, Antioxidant and antiproliferative activities of raspberries. *Journal of Agricultural and Food Chemistry*, **2003**, *50*, 2926-2930. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [27]. P. Goyal, A. Chauhan, P. Kaushik, Laboratory evaluation of crude extracts of *Cinnamomum tamala* for potential antibacterial activity, *Electronic Journal of Biology*, **2009**, *5*, 75-79. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [28]. M.S. Aliyu, U.A. Hanwa, M.B. Tijjani, A.B. Aliyu, B. Ya'u, Phytochemical and antibacterial properties of Leaf Extract of *Stereospermum kunthianum* (Bignoniaceae). *Nigerian Journal of Basic and Applied Science*, **2009**, *17*, 234-239. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [29]. V.K. Gupta, R. Gaur, A. Sharma, J. Akther, M. Saini, R.S. Bhakuni, R. Pathania, A novel bifunctional chalcone inhibits multi-drug resistant *Staphylococcus aureus* and potentiates the activity of fluoroquinolones, *Bioorganic Chemistry*, **2019**, *83*, 214-225. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [30]. M. Ayaz, F. Ullah, A. Sadiq, F. Ullah, M. Ovais, J. Ahmed, H.P. Devkota, Synergistic interactions of phytochemicals with antimicrobial agents: Potential strategy to counteract drug resistance, *Chemico-Biological Interactions*, **2019**, *308*, 294-303. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [31]. I.O. Igwenyi, C.E. Offor, D.A. Ajah, O.C. Nwankwo, J.I. Ukaomah, P.M. Aja, Chemical compositions of *Ipomea aquatica* (Green Kangkong), *International Journal of Pharmacy and Biology*, **2011**, *4*, 594-598. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [32]. R. Verpoorte, Pharmacognosy in the new millennium: Lead finding and biotechnology, *Journal of Pharmacy and Pharmacology*, **2000**, *52*, 253-262. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [33]. S.R. Kaundal, A. Sharma, R. Kumar, V. Kumar, R. Kumar, Exploration of medicinal importance of an underutilized legume crop, *Macrotyloma uniflorum* (Lam.) Verd C (Horse gram): A Review, *International Journal of Pharmaceutical Sciences Research*, **2019**, *10*, 3178-3186. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [34]. G.H. Bonjar, A.K. Nik, S. Aghighi, Antibacterial and antifungal survey in plants used in indigenous herbal-medicine of South East regions of Iran, *Journal of Biological Sciences*, **2004**, *4*, 405-412. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [35]. M.Y. Abdo, W.Y.W. Ahmad, L. bin Din, N. Ibrahim, Phytochemical study of *Hedychium malayanum* (Zingiberaceae), *Sains Malaysiana*, **2017**, *1*, 83-89. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [36]. D. Krishnaiah, T. Devi, A. Bono, R. Sarbatly, Studies on phytochemical constituent of six Malaysian medicinal plants, *Journal of Medicinal Plant Research*, **2009**, *3*, 67-72. [[Google Scholar](#)], [[Publisher](#)]
- [37]. A. Ahmad, A.F.M. Alkarkhi, S. Henaand, L.H. Lima, Extraction, separation and identification of chemical ingredients of *Elephantopus scaber* L using factorial design of experiment, *International Journal of Chemistry*, **2009**, *1*, 36-49. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [38]. S.L. Hwang, A preliminary report on the chemical composition of yam bean, (*Pachyrhizus erosus* Urban). A new rotenone bearing plant. *Kwangsi Agriculture*, **1941**, *2*, 269-280. [[Google Scholar](#)], [[Publisher](#)]
- [39]. E. Be'jar, R. Reyes-Chilpa, M. Jimenez-Estrada, Bioactive compounds from selected plants used in the XVI century Mexican traditional medicine. In: Studies in natural products chemistry, *Studies in Natural Products Chemistry*, **2000**, *24*, 799-844. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [40]. S. Alok, S.K. Jain, A. Verma, M. Kumar, Pharmacognostic and phytochemical evaluation of *Dolichos biflorus* Linn. *Asian Pacific Journal of Tropical Disease*, **2011**, *4*, S97-S101. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [41]. S.E. Jin, Y.K. Son, B.S. Min, H.A. Jung, J.S.X. Choi, Anti-inflammatory and antioxidant

- activities of constituents isolated from *Pueraria lobata* roots, *Archives of Pharmacal Research*, **2016**, *5*, 823–837. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [42]. L.V. Puyvelde, N.D. Kimpe, J.-P. Mudaheranwa, A. Gasiga, N. Schamp, J.-P. Declercq, M.V. Meerssche, Isolation and structural elucidation of potentially insecticidal *Pakistan Journal of Pharmaceutical Sciences*, **2020**, *33*, 361–369. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [44]. F.J. Álvarez-Martinez, E. Barrajon-Catalan, J.A. Encinar, J.C. Rodriguez-Diaz, V. Micol, Antimicrobial capacity of plant polyphenols against Gram-positive bacteria: A comprehensive review, *Current Medicinal Chemistry*, **2020**, *27*, 2576-2606. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [45]. A.E. Al-Snafi, Antimicrobial effects of medicinal plants (part 3): Plant Based Review, *IOSR Journal of Pharmacy*, **2016**, *6*, 67-92. [[Google Scholar](#)], [[Publisher](#)]
- [46]. M.A. Abushaheen, A.J. Fatani, M. Alosaimi, W. Mansy, M. George, S. Acharya, S. Rathod, D.D. Divakar, C. Jhugroo, S. Vellappally, A.A. Khan, Antimicrobial resistance, mechanisms and its clinical significance. *Disease-a-Month*, **2020**, *66*, 100971. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [47]. M.M. El-Zayat, M.M. Eraqi, F.A. Alfaiz, M.M. Elshaer, Antibacterial and antioxidant potential of some Egyptian medicinal plants used in traditional medicine, *Journal of King Saud University-Science*, **2021**, *33*, 101466. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [48]. P. Molyneux, The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity, *Songklanakarinn Journal of Science and Technology (SJST)*, **2004**, *26*, 211-219. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [49]. M. M. Abdel-Gawad, M.M. Abdel-Aziz, M.M. El-Sayed, E.A. El-Wakil, E.E. Abdel-Lateef, *In vitro* antioxidant, total phenolic and flavonoid contents of six *Allium* species growing in Egypt, *Journal of Microbiology, Biotechnology and Food* and acaricidal isoflavone-type compounds from *Neorautanenia mitis*, *Journal of Natural Products*, **1987**, *50*, 349–356. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [43]. M. Ambreen, S.A. Mirza, Evaluation of anti-inflammatory and wound healing potential of tannins isolated from leaf callus cultures of *Achyranthes aspera* and *Ocimum basilicum*. *Science*, **2014**, *3*, 343-346. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [50]. L.V. Puyvelde, N. De-Kimpe, J. Mudaheranwa, A. Gasiga, N. Schamp, J. Declercq, M.V. Meerssche, Isolation and structural elucidation of potentially insecticidal and acaricidal isoflavone-type compounds from *Neorautanenia mitis*, *Journal of Natural Products*, **1987**, *50*, 349-356. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [51]. M.A. Orabi, H. Aoyama, T. Kuroda, T. Hatano, Structures of two new flavonoids and effects of licorice phenolics on vancomycin-resistant *Enterococcus* species. *Molecules*, **2014**, *19*, 3883–3897. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [52]. J.P. Dzoyem, R. Melong, A.T. Tsamo, A.T. Tchinda, D.G. Kapche, B.T. Ngadjui, J.N. Eloff, Cytotoxicity, antimicrobial and antioxidant activity of eight compounds isolated from *Entada abyssinica* (Fabaceae). *BMC Research Notes*, **2017**, *10*, 118. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [53]. N.F. Shamsudin, Q.U. Ahmed, S. Mahmood, S.A. Ali Shah, A. Khatib, S. Mukhtar, M.A. Alsharif, H. Parveen, Z.A. Zakaria, Antibacterial effects of flavonoids and their structure-activity relationship study: A comparative interpretation, *Molecules*, **2022**, *27*, 1149. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [54]. A.M. Mohamed, N.M. Metwally, S.S. Mahmoud, *Sativa* seeds against *Schistosoma mansoni* different stages, *Memórias do Instituto Oswaldo Cruz*, **2005**, *100*, 205-211. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [55]. L. Cromble, D.A. Whiting, The constitution of neotenone and dolichone: biogenetic connection in the sub-family Papiilonaceae.



- Tetrahedron Letters*, **1962**, *18*, 801–804. [Crossref], [Google Scholar], [Publisher]
- [56]. A.J. Brink, Epidemiology of carbapenem-resistant Gram-negative infections, *Current Opinion in Infectious Diseases*, **2019**, *32*, 610–616. [Crossref], [Google Scholar], [Publisher]
- [57]. L. Aires Ade, E.C. Ximenes, R.A. Silva, Ultrastructural analysis of  $\beta$ -lapachone-induced *epidermidis*, *BMC Pharmacology and Toxicology*, **2016**, *17*, 39. [Crossref], [Google Scholar], [Publisher]
- [59]. R. Nanta, R.K. Kale, Chemomodulatory effect of *Dolichos biflorus* Linn. on skin and forestomach papillomagenesis in Swiss albino mice, *Indian Journal of Experimental Biology*, **2011**, *49*, 483–490. [Crossref], [Google Scholar], [Publisher]
- [60]. D. Chattopadhyay, K. Maiti, A.P. Kundu, M.S. Chakraborty, R. Bhadra, S.C. Maudal, A.B. Maudal, Antimicrobial activity of *Alstonia macrophylla*: A folklore of bay islands, *Journal of Ethnopharmacology*, **2001**, *77*, 49–55. [Crossref], [Google Scholar], [Publisher]
- [61]. W. Bahaa, E.I. Deen, G.S. Sadek, Parasitological, pathological and immunological effects of hesperidin treatment on murine schistosomiasis mansoni, *Life Science Journal*, **2014**, *11*, 840–855. [Crossref], [Google Scholar], [Publisher]
- [62]. R. Das, D. Mukherjee, S. Reja, K. Sarkar, A. Kejriwal, Copper based *N,N*-dimethyl-*N*-(1-pyridinylmethylidene) propane-1,3-diamine compound: Synthesis, characterization, and its application toward biocidal activity, *Journal of Applied Organometallic Chemistry*, **2023**, *3*, 73–85. [Crossref], [Google Scholar], [Publisher]
- [63]. S. Bano, K. Javed, S. Ahmad, I.G. Rathish, S. Singh, M. Chaitanya, K.M. Arunasree, M.S. Alama, Synthesis of some novel chalcones, flavanones and flavones and evaluation of their anti-inflammatory activity, *European Journal of Medicinal Chemistry*, **2017**, *65*, 51–59. [Crossref], [Google Scholar], [Publisher]
- [64]. R. Maharjan, S. Thapa, A. Acharya, Evaluation of antimicrobial activity and surface membrane damage in male adult *Schistosoma mansoni* BH strain worms, *Experimental Parasitology*, **2021**, *142*, 83–90. [Crossref], [Google Scholar], [Publisher]
- [58]. S. Siritwong, Y. Teethaisong, K. Thumanu, B. Dunkhunthod, G. Eumkeb, The synergy and mode of action of quercetin plus amoxicillin against amoxicillin-resistant *Staphylococcus* synergistic effect of spices against few selected pathogens, *Tribhuvan University Journal of Microbiology*, **2019**, *6*, 10–18. [Crossref], [Google Scholar], [Publisher]
- [65]. A.G.V. Costa, D.F. Garcia-Diaz, P. Jimenez, P.I. Silva, Bioactive compounds and health benefits of exotic tropical red-black berries, *Journal of Functional Foods*, **2020**, *5*, 539–549. [Crossref], [Google Scholar], [Publisher]
- [66]. J. de Moraes, Natural products with antischistosomal activity review, *Future Medicinal Chemistry*, **2015**, *7*, 801–820. [Crossref], [Google Scholar], [Publisher]
- [67]. SH. Xiao, B.A. Catto, *In vitro* and *in vivo* studies of the effect of artemether on *Schistosoma mansoni*, *Antimicrobial agents and chemotherapy*, **1989**, *33*, 1557–1562. [Crossref], [Google Scholar], [Publisher]
- [68]. J. Boissier, F. Cosledan, A. Robert, *In vitro* activities of trioxaquinones against *Schistosoma mansoni*, *Antimicrobial agents and chemotherapy*, **2009**, *53*, 4903–4906. [Crossref], [Google Scholar], [Publisher]
- [69]. Y.X. Liu, W. Wu, Y.J. Liang, New uses for old drugs: the tale of artemisinin derivatives in the elimination of schistosomiasis japonica in China, *Molecules*, **2014**, *19*, 15058–15074. [Crossref], [Google Scholar], [Publisher]
- [70]. F. Farhadi, B. Khameneh, M. Iranshahi, M. Iranshahi, Antibacterial activity of flavonoids and their structure-activity relationship: An update review, *Phytotherapy Research*, **2019**, *33*, 13–40. [Crossref], [Google Scholar], [Publisher]
- [71]. J. de Moraes, A.A.C. Almeida, M.R.M. Brito, Anthelmintic activity of the natural compound (+)-limonene epoxide against *Schistosoma*

- mansoni*, *Planta Medica*, **2013**, *79*, 253–258. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [72]. J. Utzinger, S. Xiao, E.K. N'Goran, The potential of artemether for the control of schistosomiasis, *International Journal for Parasitology*, **2001**, *31*, 1549–1562. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [73]. W. Jiraungkoorskul, S. Sahaphong, P. Sobhon, *Schistosoma mekongi*: the *in vitro* effect of praziquantel and artesunate on the adult fluke, *Experimental Parasitology*, **2006**, *113*, 16–23. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [74]. B. Jubeh, Z. Breijyeh, R. Karaman, Resistance of gram-positive bacteria to current antibacterial agents and overcoming approaches, *Molecules*, **2020**, *25*, 2888. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [75]. H. Sangho, A. Dabo, H. Coulibaly, O. Doumbo, Prevalence and perception of schistosomiasis in peri urban schools of Bamako in Mali. *Bulletin de la Soci'ete de Pathologie Exotique*, **2002**, *95*, 292–294. [[Google Scholar](#)], [[Publisher](#)]
- [76]. H.J. Li, W. Wang, G.L. Qu, Effect of the *in vivo* activity of dihydroartemisinin against *Schistosoma mansoni* infection in mice, *Parasitology Research*, **2012**, *110*, 1727–1732. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [77]. M.P.N. Silva, G.L.S. Oliveira, R.B.F. de Carvalho, Antischistosomal activity of the terpene nerolidol. *Molecules*, **2014**, *19*, 3793–3803. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [78]. D.C. Sass, G.O. Morais, R.A. Miranda, Structurally modified natural sesquiterpene lactones constitute effective and less toxic schistosomicidal compounds, *Organic & Biomolecular Chemistry*, **2014**, *12*, 7957–7964. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [79]. J. de Moraes, R.N. de Oliveira, J.P. Costa, Phytol, a diterpene alcohol from chlorophyll, as a drug against neglected tropical disease Schistosomiasis mansoni, *PLOS Neglected Tropical Diseases*, **2014**, *8*, e2617. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [80]. N.M. Musa, M.S. Sallau, O.A. Oyewale, T. Ali, Isolation and characterization of neoduline from the rhizome of *Dolichos pachyrhizus* harm, *Advanced Journal of Chemistry – Section B*, **2022**, *4*, 113–123. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [81]. A.Y. Spivak, J. Keiser, M. Vargas, Synthesis and activity of new triphenylphosphonium derivatives of betulin and betulinic acid against *Schistosoma mansoni* *in vitro* and *in vivo*, *Bioorganic & Medicinal Chemistry*, **2014**, *22*, 6297–6304. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [82]. C. Ramalheite, L.G. Magalhaes, V. Rodrigues, *In vitro* schistosomicidal activity of balsaminol F and karavilagenin C, *Planta Medica*, **2012**, *78*, 1912–1917. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [83]. J. de Moraes, C. Nascimento, L.F. Yamaguchi, *Schistosoma mansoni*: *In vitro* schistosomicidal activity and tegumental alterations induced by pipartine on schistosomula, *Experimental Parasitology*, **2012**, *132*, 222–227. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [84]. V.S. Carrara, S.C. Vieira, R.G. de Paula, *In vitro* schistosomicidal effects of aqueous and dichloromethane fractions from leaves and stems of *Piper* species and the isolation of an active amide from *P. amalago* L. (Piperaceae), *Journal of Helminthology*, **2014**, *88*, 321–326. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [85]. L.M. Veras, M.A. Guimaraes, Y.D. Campelo, Activity of episopiloturine against *Schistosoma mansoni*, *Current Medicinal Chemistry*, **2012**, *19*, 2051–2058. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [86]. M.A. Miranda, L.G. Magalhaes, R.F. Tiossi, Evaluation of the schistosomicidal activity of the steroidal alkaloids from *Solanum lycocarpum* fruits, *Parasitology Research*, **2012**, *111*, 257–262. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [87]. J. Keiser, J. Chollet, S.H. Xiao, Mefloquine-an aminoalcohol with promising antischistosomal properties in mice, *PLOS*

- Neglected Tropical Diseases*, **2009**, *3*, e350. [Crossref], [Google Scholar], [Publisher]
- [88]. S.H. Xiao, J.Y. Mei, P.Y. Jiao, The *in vitro* effect of mefloquine and praziquantel against juvenile and adult *Schistosoma japonicum*. *Parasitology Research*, **2009**, *106*, 237–246. [Crossref], [Google Scholar], [Publisher]
- [89]. S.M. Zhang, K.A. Coultas, Identification of plumbagin and sanguinarine as effective *Experimental Parasitology*, **2013**, *133*, 18–27. [Crossref], [Google Scholar], [Publisher]
- [91]. L. Aires Ade, E.C. Ximenes, V.X. Barbosa,  $\beta$ -Lapachone: a naphthoquinone with promising antischistosomal properties in mice, *Phytomedicine*, **2014**, *21*, 261–267. [Crossref], [Google Scholar], [Publisher]
- [92]. N.L. Cunha, C.J. Uchoa, L.S. Cintra, *In vitro* schistosomicidal activity of some Brazilian cerrado species and their isolated compounds, *Evidence-Based Complementary and Alternative Medicine*, **2012**, *2012*, 173614 [Crossref], [Google Scholar], [Publisher]
- [93]. C.G. Braguine, C.S. Bertanha, U.O. Goncalves, Schistosomicidal evaluation of flavonoids from two species of *Styrax* against *Schistosoma mansoni* adult worms, *Pharmaceutical Biology*, **2012**, *50*, 925–929. [Crossref], [Google Scholar], [Publisher]
- [94]. C.J. Xiao, Y. Zhang, L. Qiu, Schistosomicidal and antioxidant flavonoids from *Astragalus englerianus*, *Planta Medica*, **2014**, *80*, 1727–1731. [Crossref], [Google Scholar], [Publisher]
- [95]. G. Allam, A.S. Abuelsaad, *In vitro* and *in vivo* effects of hesperidin treatment on adult worms of *Schistosoma mansoni*, *Journal of Helminthology*, **2014**, *88*, 362–370. [Google Scholar], [Publisher]
- [96]. A.C. Pereira, L.G. Magalhaes, U.O. Goncalves, Schistosomicidal and trypanocidal structureactivity relationships for ( $\pm$ )-licarin A and its (-)- and (+)-enantiomers. chemotherapeutic agents for treatment of schistosomiasis, *International Journal for Parasitology: Drugs and Drug Resistance*, **2013**, *3*, 28–34. [Crossref], [Google Scholar], [Publisher]
- [90]. N. Lorsuwannarat, N. Saowakon, P. Ramasoota, The anthelmintic effect of plumbagin on *Schistosoma mansoni*, *Phytochemistry*, **2011**, *72*, 1424–1430. [Crossref], [Google Scholar], [Publisher]
- [97]. A.K. El-Ansary, S.A. Ahmed, S.A. Aly, Antischistosomal and liver protective effects of *Curcuma longa* extract in *Schistosoma mansoni* infected mice, *Indian Journal of Experimental Biology (IJEB)*, **2007**, *45*, 791–801. [Crossref], [Google Scholar], [Publisher]
- [98]. M.A. El-Banhawey, M.A. Ashry, A.K. El-Ansary, Effect of curcuma longa or parziquantel on *Schistosoma mansoni* infected mice liver-histological and histochemical study, *Indian Journal of Experimental Biology (IJEB)*, **2007**, *45*, 877–889. [Crossref], [Google Scholar], [Publisher]
- [99]. L.G. Magalhaes, G.J. Kapadia, L.R. da Silva Tonuci, *In vitro* schistosomicidal effects of some phloroglucinol derivatives from *Dryopteris* species against *Schistosoma mansoni* adult worms, *Parasitology Research*, **2010**, *106*, 395–401. [Crossref], [Google Scholar], [Publisher]
- [100]. Y.Q. Chen, Q.M. Xu, X.R. Li, *In vitro* evaluation of schistosomicidal potential of curcumin against *Schistosoma japonicum*, *Journal of Asian Natural Products Research*, **2012**, *14*, 1064–1072. [Crossref], [Google Scholar], [Publisher]
- [101]. L.G. Magalhaes, C.B. Machado, E.R. Morais, *In vitro* schistosomicidal activity of curcumin against *Schistosoma mansoni* adult worms, *Parasitology Research*, **2009**, *104*, 1197–1201. [Crossref], [Google Scholar], [Publisher]